Beiwudine, a Norditerpenoid Alkaloid from Aconitum kusnezoffii

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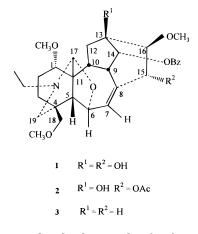
A novel norditerpenoid alkaloid, beiwudine (1), was isolated from the roots of *Aconitum kusnezoffii*. Its structure was established on the basis of chemical and NMR spectral studies.

The root part of *Aconitum kusnezoffli* Reichb. (Ranunculaceae), native to northern China, is used in folk medicine for the treatment of rheumatism and neuralgia.¹ In the previous papers,^{2–4} a number of norditerpenoid alkaloids and diterpenoid alkaloids were reported from this plant. Continued investigation of this plant has now led to the isolation of a novel norditerpenoid alkaloid, beiwudine (1). This paper deals with the isolation and structure elucidation of the compound.

Compound 1 was isolated as an amorphous powder. The HREIMS showed a molecular ion $[M]^+$ at m/z 555.2953 consistent with the molecular formula $C_{31}H_{41}NO_8$. The ¹H and ¹³C NMR spectra indicated the presence of an N-ethyl $[\delta_{\rm H} 1.00 \text{ (3H, t, } J = 7.2 \text{ Hz}); \delta_{\rm C} 13.0 \text{ q}, 49.1 \text{ t}], \text{ three methoxy}$ groups ($\delta_{\rm H}$, 3.26, 3.30, 3.67; $\delta_{\rm C}$ 57.1, 59.4, 61.3), and a benzoyl ester group [$\delta_{\rm H}$ 7.45 (2H, t, J = 7.2 Hz), 7.57 (1H, t, J = 7.2 Hz), 8.07 (2H, d, J = 7 Hz); $\delta_{\rm C}$ see Table 1]. The spectral data of 1 were quite similar to those of franchetine (3),⁵ and the compound was considered to possess a norditerpenoid alkaloid structure. Thus, examination of the ¹H-¹H and ¹H-¹³C COSY spectra led to the unambiguous assignments of C-1 through C- 3 signals; and the chemical shift of C-1 (δ 86.3) clearly indicated the presence of a methoxyl group. Another methoxyl group (δ 59.4) was assigned to C-18 because the latter was readily identified as an oxymethylene at δ 78.9, consistent with other compounds bearing 18-OCH₃ group.^{6,7} The NMR data further suggested the presence of an N,O-mixed acetal group [$\delta_{\rm H}$ 4.37 (1H, s), 4.55 (1H, d, J = 6.4 Hz); $\delta_{\rm C}$ 92.6 d, 74.3 d]. In an NOE experiment, irradiation of the signal at δ 4.55 (H-6) enhanced both the signals for H-5 (δ 2.29, s, 3%) and 18-CH₂ (δ 3.02, 3.12, ABq, 9%), and vice versa. Hence, 1 was considered to be a franchetine derivative bearing an oxygen bridge between C-6 and C-17.

A difference of 32 mass units between **1** and franchetine (**3**) suggests that the former is a dihydroxyl derivative. A hydroxyl group was assigned to C-15 on the basis of the coupling relationship observed between H-15 (δ 5.04, d) and H-16 (δ 2.85, d). This assignment was also supported by the result of an NOE experiment on acetylbeiwudine (**2**) in which NOE was observed between H-7 (δ 6.16, d) and 15-OAc ($\delta_{\rm H}$ 2.15, s). The other OH functional group, locating on a tertiary carbon (δ 76.5), was assigned to C-13 based on the downshift of the C-16 as in other 13,15-dihydroxy derivatives.⁸

Attention was then focused on the determination of the configuration of the 15-OH group. In order to observe the NOE results, **1** was acetylated to **2**. Irradiation of H-7 (δ 5.81) led to the enhancement of the signals of 15-OAc (3.5%) and H-6 (13%); and irradiation of H-15 (δ 6.16) resulted in an enhancement of H-2"/H-6" of the benzene ring (13%). These results indicated that the 15-OAc was α -oriented.



Beiwudine is the third example of a franchetine-type norditerpenoid alkaloid bearing an *N*,*O*-acetal [N-C(17)-O-C(6)] with a C-7/C-8 double bond. Assignments of its ¹H and ¹³C NMR data were made by analysis of COSY, HETCOR, COLOC, NOESY, and NOEDS results.

Experimental Section

General Experimental Procedures. ¹H and ¹³C NMR spectra were determined in CDCl₃, with TMS as internal standard, on a Bruker ARX-300 spectrometer. LRMS data were recorded on either a Finnigan TSQ 7000 mass spectrometer or a M-80A GL–MS spectrometer. HRMS spectra were measured on a Kratos MS-80 spectrometer. A polyvinyl sulfonic ion resin (H form, cross linking 1×3 , Chemical Factory of Nan Kai University, China) was used in the extraction of total alkaloids. Column chromatography was carried out on Si gel H, and TLC on Si gel G plates, with solvent systems S₁(CHCl₃–MeOH, 9:1) and S₂ (ether–Me₂CO, 9:1), detected with modified Dragendorffs reagent or I₂ vapor. Si gels H and G were purchased from the Qingdao Marine Chemical Factory, China.

Plant Material. *A. kusnezoffii* roots were collected in September 1991, in Chifeng of Inner Mongolia. The plant was identified by Prof. W. T. Wang (Institute of Botany, Chinese Academy of Sciences, Beijing), and voucher specimens have been deposited in the herbarium of the School of Pharmacy, West China University of Medical Sciences.

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Table 1.	NMR Data	for Beiwudine	(1) and	Franchetine ((3) ⁵
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position	3 ¹³ C	1			
		¹³ C	¹ H	COLOC	NOESY
1	86.5	86.3 d	ca. 3.30 m		H-5
2	24.3	24.3 t	ca. 1.80 m	C-11	
			ca. 2.40 m		
3	32.7	32.7 t	ca. 1.50 m		
			ca. 1.70 m		H-5
4	37.3	37.3 s			
5	47.9	47.5 d	2.29 s	C-10, C-l , C-17	H-1 β , H-3 β , 18-CH ₂
6	74.8	74.3 d	4.55 d (6.4)		H-7, 18-CH ₂
7	128.7	123.9 d	6.08 d (6.4)		H-6
8	136.8	138.6 s			
9	42.9	42.7 d	ca. 3.10 m		
10	49.4	47.5 d	2.65 m	C-11	H-14
11	50.1	50.5 s			
12	29.7	38.9 t	ca. 2.00 m	C-9, C-16	H-17
13	38.3	76.5 s			
14	78.7	74.6 d	5.04 br s ($W_{1/2} = 4.6$ Hz)	C-13	H-10
15	38.5	83.3 d	5.04 d(6)		
16	85.4	94.2 d	2.85 d (overlapped)		
17	92.2	92.6 d	4.37 s	C-1, C-6	12-CH ₂
18	78.8	78.9 t	3.02, 3.25 (ABq, 9.2)		H-5, H-6
19	52.1	52.0 t	2.03, 2.35 (ABq, hidden)		
NCH2CH3	49.0	49.1 t	2.35 m, 2.55 m		
$N\overline{CH}_2CH_3$	13.0	13.0 q	1.00 t (7.1)		
$1-OCH_3$	57.6	57.1 q	3.30 s		
16-OCH ₃	55.9	61.3 q	3.67 s	C-16	
18-OCH ₃	59.3	59.4 q	3.26 s		
benzoyl ester:		-			
C=0	166.4	166.6 s			
1″	130.5	129.6 s			
2", 6"	128.8	129.5 d	8.07 d (7)	C-4″	H-15
3", 5"	128.2	128.5 d	7.45 d (7.2)		
4‴	132.6	133.2 d	7.57 d (7.2)		

Extraction and Isolation. Powdered roots of A. kusnezoffii (8 kg) were percolated with 0.15% HCl (33 L). Wet resin (dry wt 2.5 kg) was added to the percolates. After exchange, the resin was washed repeatedly on a suction filter with deionized H₂O and air dried. The dried resin was then mixed with 10% aqueous NH₄OH (total amount 1 L) and extracted in a specially designed extractor⁹ with Et₂O under reflux until no alkaloid could be detected with Dragendorff's reagent. The total alkaloid fraction so obtained (32.8 g) appeared as a white powder. The total alkaloid (8 g) was subsequently chromatographed on Si gel H (280 g) eluted with CHCl₃-MeOH (96:4) to afford fractions A (1.6 g), B (2.5 g), and C (3.5 g). Fraction A was chromatographed on Si gel H eluted with CHCl₃–MeOH (98.5:1.5 to 96:4) (10-mL fractions) to afford fractions A-1 (400 mg), A-2 (250 mg), and A-3 (1.1 g). Fraction A-1 was chromatographed on Si gel H eluting with cyclohexane-Me₂COdiethlyamine (80:20:1) to give a white amorphous powder (1, 120 mg) showing one spot on TLC (S₁, S₂). Fraction A-3 was chromatographed on Si gel H eluting with cyclohexane-Me₂CO-diethylamine (70:30:1) to give hypaconitine (117 mg) and aconifine (90 mg).4 Fraction C was chromatographed repeatly on Si gel H (100 g) eluting with CHCl3-MeOH (99:1 to 9:1) (15-mL fractions) to afford beiwutine (460 mg) and aconifine (300 mg).4

Beiwudine (1): white amorphous powder; IR (KBr) ν_{max} 3459 (OH), 1718, 1279 cm⁻¹; ¹H (300 MHz) and ¹³C (75 MHz) NMR, see in Table 1; EIMS *m*/*z* 555 ([M]⁺, 8), 540 ([M – Me]⁺, 11), 524 ([M - OMe]⁺, 44), 105 ([C₆H₅ - CO]⁺, 100); HREIMS m/z 555.2953 (calcd for C₃₁H₄₁O₈, 555.2911).

Acetylation of 1. To 1 (80 mg) was added pyridine (0.5 mL) and acetyl anhydride (0.5 mL). The solution was allowed to stand at room temperature overnight. After removal of the solvent under reduced pressure, the residue was chromatographed on a chromatotron eluting with CHCl₃-MeOH (99: 1) to give a white amorphous powder of acetylbeiwudine (2) (42 mg) showing one spot on TLC (S_1 , S_2).

Acetylbeiwudine (2): ¹H NMR (300 MHz) δ 1.01 (3H, t, J = 7.2 Hz, NCH₂*CH*₃), 2.15 (3H, s, OAc), 3.06, 3.15 (each 1H, ABq, J = 9.0 Hz, H₂-18), 3.24, 3.36, 3.51 (each 3H, s, 3 \times

OCH₃), 4.37 (1H, s, H-17), 4.51 (1H, d, *J* = 6.3 Hz, H-6), 5.13 (1H, br s, H-14 β), 5.81 (1H, d, J = 6.0 Hz, H-7), 6.16 (1H, d, J= 6.0 Hz, H-15 β), 7.56, 7.45, 8.08 (5H, m, aromatic protons); $^{13}\mathrm{C}$ NMR (75 MHz) δ 86.2 d (C-1), 24.1 t (C-2), 32.5 t (C-3), 37.2 (C-4), 47.2 d (C-5), 74.2 d (C-6), 123.9 d (C-7), 133.0 s (C-8), 42.7 d (C-9), 47.2 d (C-10), 50.4 s (C-11), 38.9 t (C-12), 76.5 s (C-13), 82.7 d (C-14), 76.0 d (C-15), 90.4 d (C-16), 92.5 d (C-17), 78.6 t (C-18), 52.0 t (C-19), 49.0 t (NCH₂CH₃), 12.9 q (NCH₂*CH*₃), 56.9 q (C-1'), 60.9 q (C-16'), 59.2 q (C-18'), 169.8 s, 20.9 q (OC-CH₃), 166.5 s (C₆H₅-CO), 129.7 s (C-1"), 129.7 d (C-2", C-6"), 128.3 d (C-3", C-5"), 133.0 d (C-4"). EIMS m/z 597 ([M]⁺, 2), 566 ([M - OCH₃]⁺, 20), 105 ([C₆H₅ - CO]⁺; HREIMS *m*/*z* 597.3010 (calcd for C₃₃H₄₃O₉N, 597.2937).

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